

1-1-2010

A general co-expression network-based approach to gene expression analysis: comparison and applications

Jianhua Ruan

University of Texas at San Antonio

Angela K. Dean

University of Texas at San Antonio

Weixiong Zhang

Washington University School of Medicine in St. Louis

Follow this and additional works at: http://digitalcommons.wustl.edu/open_access_pubs



Part of the [Medicine and Health Sciences Commons](#)

Recommended Citation

Ruan, Jianhua; Dean, Angela K.; and Zhang, Weixiong, "A general co-expression network-based approach to gene expression analysis: comparison and applications." *BMC Systems Biology*. 4, . 8. (2010).
http://digitalcommons.wustl.edu/open_access_pubs/89

This Open Access Publication is brought to you for free and open access by Digital Commons@Becker. It has been accepted for inclusion in Open Access Publications by an authorized administrator of Digital Commons@Becker. For more information, please contact engeszer@wustl.edu.

METHODOLOGY ARTICLE

Open Access

A general co-expression network-based approach to gene expression analysis: comparison and applications

Jianhua Ruan^{1*}, Angela K Dean¹, Weixiong Zhang^{2,3*}

Abstract

Background: Co-expression network-based approaches have become popular in analyzing microarray data, such as for detecting functional gene modules. However, co-expression networks are often constructed by ad hoc methods, and network-based analyses have not been shown to outperform the conventional cluster analyses, partially due to the lack of an unbiased evaluation metric.

Results: Here, we develop a general co-expression network-based approach for analyzing both genes and samples in microarray data. Our approach consists of a simple but robust rank-based network construction method, a parameter-free module discovery algorithm and a novel reference network-based metric for module evaluation. We report some interesting topological properties of rank-based co-expression networks that are very different from that of value-based networks in the literature. Using a large set of synthetic and real microarray data, we demonstrate the superior performance of our approach over several popular existing algorithms. Applications of our approach to yeast, Arabidopsis and human cancer microarray data reveal many interesting modules, including a fatal subtype of lymphoma and a gene module regulating yeast telomere integrity, which were missed by the existing methods.

Conclusions: We demonstrated that our novel approach is very effective in discovering the modular structures in microarray data, both for genes and for samples. As the method is essentially parameter-free, it may be applied to large data sets where the number of clusters is difficult to estimate. The method is also very general and can be applied to other types of data. A MATLAB implementation of our algorithm can be downloaded from <http://cs.utsa.edu/~jruan/Software.html>.

Background

The vast amount of available high-throughput gene expression data has provided excellent opportunities for studying gene functions on a global scale. Since genes on the same pathways or in the same functional complex often exhibit similar expression patterns under diverse temporal and physiological conditions, one common practice in microarray data analysis is to cluster genes according to their expression similarities. The clusters can then be analyzed in several ways, such as promoter analysis or gene ontology analysis [1]. Since

clustering procedures are often subjective, and usually ignore the detailed relationships among genes, the biological insight obtained from clustering results is often limited. Alternatively, many studies have attempted to construct gene regulatory networks from microarray data using methods such as linear models [2], Bayesian networks [3] and Boolean networks [4]. These approaches, however, are successful only on rare cases where the system is well constrained and the training data is sufficiently large.

Recently, as a tradeoff between the crude cluster analysis and detailed network modeling, gene co-expression networks have become a rapidly developing area of study and many interesting results have been obtained [5-20]. A gene co-expression network is an undirected graph, where the graph nodes correspond to genes, and

* Correspondence: jruan@cs.utsa.edu; weixiong.zhang@wustl.edu

¹Department of Computer Science, The University of Texas at San Antonio, One UTSA Circle, San Antonio, TX 78249, USA

²Department of Computer Science and Engineering, Washington University in St Louis, One Brookings Dr, St Louis, MO 63130, USA

edges between genes represent significant co-expression relationships [10,18]. Compared to regulatory networks, a gene co-expression network does not attempt to distinguish direct gene interactions from indirect ones; on the other hand, a gene co-expression network contains gene neighborhood relations that are usually overlooked in cluster analysis [21]. gives an interesting geometric interpretation of gene co-expression networks and connects co-expression network analysis to traditional microarray data analysis techniques. Databases of gene co-expression networks for several model organisms have been constructed from large numbers of microarray data sets [22,23].

One of the most important applications of gene co-expression networks is to identify functional gene modules, which often manifest themselves as dense subnetworks. While earlier studies attempted to apply clustering algorithms directly to the adjacency matrices of networks in order to partition network nodes into groups [10,14], later studies have relied on graph partitioning algorithms or special purpose algorithms for identifying subnetworks of certain properties [5-7,11,17,18,20]. These studies have provided many interesting biological results; however, none of them has demonstrated that the network-based methods for detecting functional modules can significantly outperform the conventional cluster analysis, partially due to the lack of an unbiased metric for quantitatively evaluating the functional significance of gene modules. In principle, nearly all the previous gene co-expression network construction methods fall into two categories: those that utilize the similarity values (value-based) [5-16,24], and those that utilize the rank-transformed similarities (rank-based) [17,18,20]. In the value-based (rank-based) method, two genes are connected if the (rank-transformed) similarity between their expression profiles is above a certain threshold, which can be chosen in an ad hoc manner or semi-automatically. Similarities may be measured by Pearson correlation coefficient or other metrics. Some researchers proposed to use conditional correlations to partially remove indirect links [24]. Thresholds are usually chosen using an ad hoc methods [16], or by controlling statistical significance of similarities [10,19]. A semi-automatic method has also been proposed based the topological difference between real and random co-expression networks [8]. In addition, several methods have been proposed to adjust raw similarity values based on neighborhood information such as shortest path connections or topological overlaps, which usually result in weighted gene co-expression networks [6,16]. Most of the existing studies used the value-based method, while the relative advantages and disadvantages of the rank-based or the value-based network construction methods have not been rigorously examined.

A number of studies have analyzed the topological properties of gene co-expression networks [10-13,18,25], and have shown that gene co-expression networks have the well-known small-world and scale-free properties, similar to many other biological networks and real-world networks [26,27]. On the other hand, these studies have also reported that gene co-expression networks differ from other types of biological networks in several important aspects, such as the characteristic node degree and hierarchical organization. However, most of the conclusions were drawn from value-based co-expression networks. In fact, we will show that the rank-based network has all the common topological properties of the other biological networks, and is significantly different from the value-based network.

In this work, we propose a general co-expression network-based approach for analyzing microarray data, for both genes and samples. Our approach consists of a method for rank-based network construction, a parameter-free graph partitioning algorithm for module discovery and a novel reference network-based metric for module evaluation. We compare our approach with the existing methods, and show several real applications of our approach.

We first introduce a simple rank-based method to construct co-expression networks, and compare the topologies of value-based and rank-based gene co-expression networks. We find that the rank-based network significantly differs from the value-based network in several important aspects, and we argue that the former is able to better capture the global topology of the underlying biological system, including both strongly and weakly co-expressed modules, while the latter is dominated by the most strongly co-expressed modules.

Second, we introduce an efficient graph partitioning algorithm, *Qcut*, to identify relatively dense modules, also known as communities, in the rank-based networks. Compared to other graph partitioning algorithms, *Qcut* is parameter free, as it uses an objective function called modularity to automatically determine the optimal partitioning and the number of partitions [28]. We have recently shown that the method can be used to discover natural communities from social networks and to predict protein complexes from protein-protein interaction networks [28], while its effectiveness in finding intrinsic modular structures from gene expression data has not been thoroughly studied.

Third, we propose a novel metric for comparing different module detection algorithms in terms of the overall functional significance of the gene modules being identified. We introduce the concept of a reference network, which can be obtained from other information sources, such as protein-protein interaction networks or ChIP-chip data. This metric is then used to quantify the

performance of a gene module detection algorithm by the agreement between the modules identified from the co-expression network and the modular structure of the reference network. It is important to note this metric is not biased by the number of modules and the module size distribution.

We assessed our approach for finding gene modules on a large set of synthetic microarray data with known modular structures as well as several real data sets. On synthetic data, our method correctly predicted the number of modules, and performed significantly better than several popular clustering algorithms and one of the best graph partitioning algorithms, the Markov Clustering algorithm [29]. On real data, we evaluated the performance of our method using both a popular (but in our opinion biased) metric based on gene ontology enrichment scores, and the aforementioned reference network-based metric. We show that our method can significantly outperform the existing algorithms according to all the metrics used.

We also applied our method to construct and analyze a sample co-expression network using microarray data of normal and cancerous T and B cells including diffuse large B-cell lymphoma (DLBCL) [30]. Remarkably, our method almost perfectly separated the different cell types into their own subnetworks without any prior knowledge. We also re-discovered the two known subtypes of DLBCL, where one subtype has a much lower survival rate than the other.

Finally, we report several interesting results that may worth further investigation. In yeast, we identified many gene modules that are both strongly co-expressed and co-regulated; among them, a small module where majority genes have no known functions may be involved in telomere maintenance. In the sample co-expression network, we discovered a tumor cell module that seems to be a new subtype of DLBCL that is associated with the lowest survival rate among all DLBCL patients.

Results and Discussion

Co-expression network construction and topological analysis

Constructing co-expression networks

Most of the existing co-expression analyses construct value-based networks. We believe that the value-based methods are significantly limited by their use of a homogeneous threshold for all the genes in the network. In reality, genes in different functional pathways may be regulated by different mechanisms, and therefore may exhibit different patterns of co-expression. In particular, genes in one functional pathway may be strongly mutually co-expressed, while genes in another functional pathway may be only weakly co-expressed. As a result, if we choose a stringent global threshold, many genes in

the weakly co-expressed pathway may be disconnected. On the other hand, if we attempt to connect the weakly co-expressed genes into the network, the threshold may become so low that the genes in the strongly co-expressed pathway may have many links to genes in other pathways, making further analysis difficult. For example, as shown in Figure 1, to construct a co-expression network for the 3000 yeast genes that we will see in the next subsection, if we allow only 10% of the genes to have no connections, most genes will have more than 300 connections, while if we reduce the median degree to 10, more than one third of the genes will have no connection at all.

To deal with this problem, we propose a simple rank-based method to construct co-expression networks. We first calculate the Pearson correlation coefficient (or some other similarity measure) between every pair of genes. For each gene g_i , we rank all other genes by their similarity to g_i . We then connect every gene to the d genes that are most similar to it. Compared to the value-based method, the rank-based method essentially uses different local similarity threshold for different genes. It is important to mention that even with a fixed d , the number of connections for different genes is not constant. This is because of the asymmetric nature of the ranking. In other words, the rank of gene i with respect to gene j is not necessarily equal to the rank of gene j with respect to gene i . Therefore, although gene i has only d genes on its top- d list, other genes that are not on i 's list may list i as one of their top- d genes. The mean degree is between d and $2d$, the minimum degree is d , and the maximum degree can be as large as $n - 1$, with n being the number of genes in the network.

The rank-based method may appear to be limited by a similar drawback of the value-based method - the former uses a global rank threshold and the latter uses a global value threshold for all genes. However, as we discussed above, in the rank-based network, different genes can have different number of connections, even though all genes have the same rank threshold, because of the asymmetric nature of ranking. More importantly, our objective is not to identify *all* co-expressed genes for each gene, but to construct a sparse network such that the modular structure of the system can be successfully identified. To achieve this, a good co-expression network needs to have the following two properties: (i) there are very few false-positive connections, and (ii) nodes within modules are well connected into a single component, while connections between modules are sparse. A value-based network can hardly provide the two properties simultaneously, for reasons given above. In contrast, the key idea in our rank-based method is that by using a uniform small value of d , we ensure that (i) the network only contains highly reliable edges, and

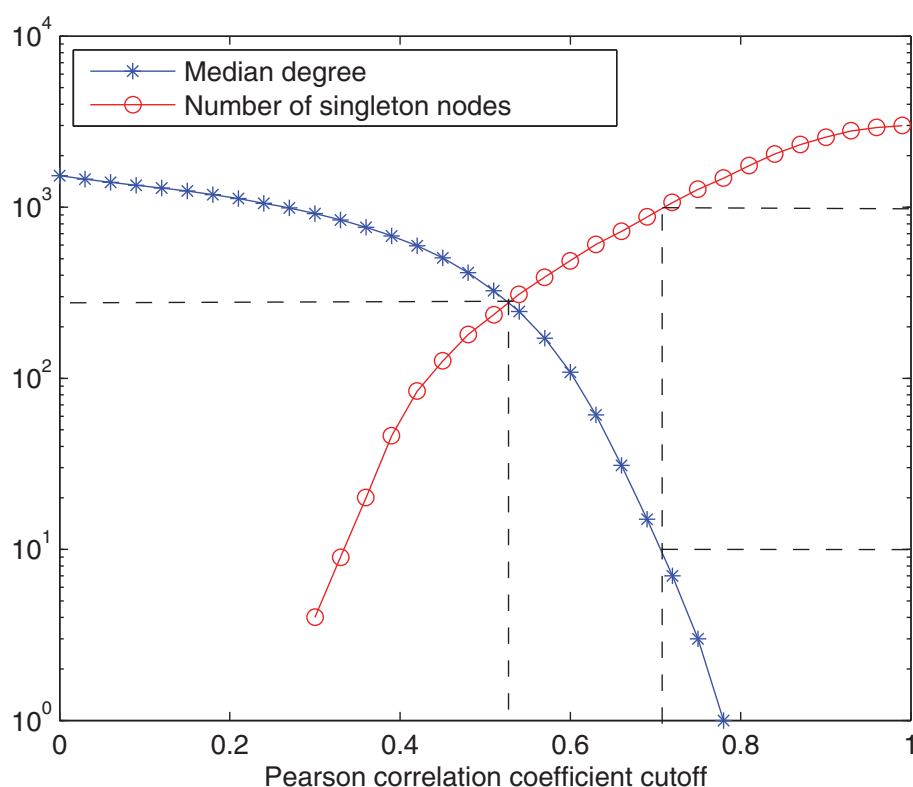


Figure 1 Median degree and number of singleton nodes in a value-based yeast co-expression network. Horizontal axis: the Pearson correlation coefficient threshold for the value-based network construction. Left vertical axis: median number of co-expression links per gene. Right vertical axis: number of genes without a co-expression link.

(ii) each module of the network is (almost) fully connected into a single component, both theoretically and empirically (see below). As in most clustering algorithms, we assume that gene expressions in different modules are generated by different distributions, while gene expressions in the same module are generated by a common (unknown) distribution. Therefore, the rank-based sub-network of genes from the same module is a nearest neighbor graph constructed on a set of random geometric points. Theoretically, it is known that a nearest neighbor graph on random geometric points has a high probability to be connected even with a very small number of neighbors (d) [31]. To empirically test this as well as to find the range of d for typical microarray data, we randomly generated a data set with 1000 genes and various dimensions (conditions) using Gaussian distribution. We then constructed a rank-based co-expression network using different values of d and measured the number of disconnected components in the resulting network. Remarkably, we find that for data of dimension > 10 , a nearest neighbor graph with 1000 nodes is almost always fully connected with $d = 2$ neighbors. Even for data of smaller dimensions, the graph can be connected with at most 4 neighbors (Figure 2). The

results do not vary significantly when the number of genes or the type of distribution is changed. In the next subsection, we also show that the yeast gene co-expression network is connected with $d = 2$. In practice we find a value of d between 3 and 5 is sufficient for most cases. This simple network construction method can also be combined with other strategies that were developed for value-based networks. For example, the raw similarity values can be refined by considering local neighborhood or shortest path information before rank transformation [16]; when selecting edges according to ranks, a threshold based on raw similarity values may be imposed simultaneously to ensure confidence in the edges being created. Ideally, methods can also be developed to automatically select the optimal d , as in [8]. The rank-based method can also be applied to construct networks of other entities, as long as a similarity measure can be defined. One example is to construct a network of samples from microarray data, where the nodes are samples and the similarity between two samples can be measured by the Pearson correlation coefficient between their gene expression profiles. Later we will show an application of a sample co-expression network where each sample is a cell type.

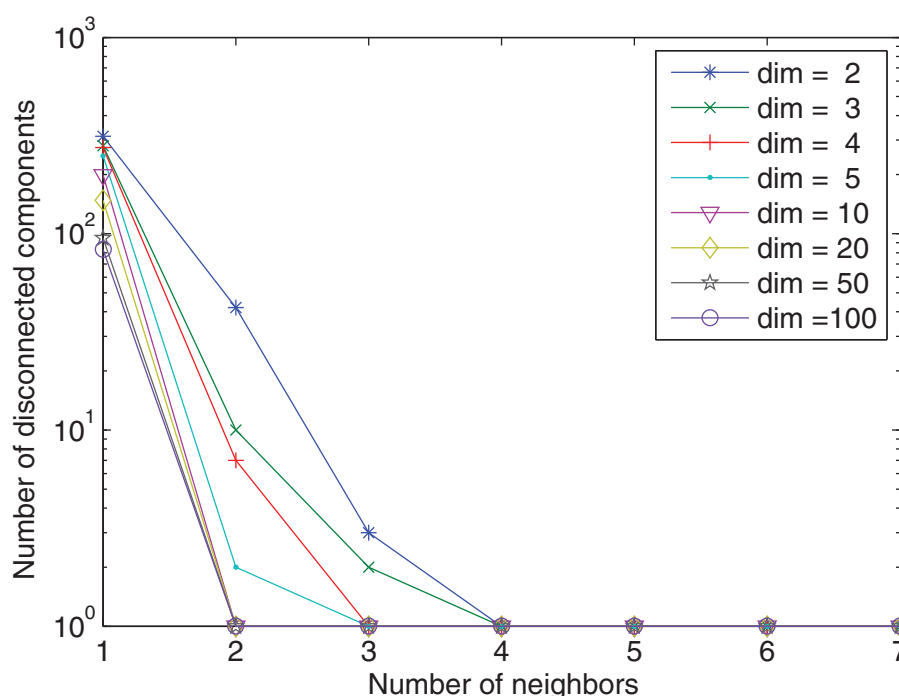


Figure 2 Connectivity of rank-based co-expression networks on random data. Each data set contains 1000 random geometric points in a certain number of dimensions, generated using the standard Gaussian distribution. Y-axis shows the number of disconnected components in the co-expression network constructed by the rank-based approach.

Topology of yeast co-expression networks

Previous studies have analyzed the topologies of various networks, including biological and social networks, and suggested three common topological properties: scale-free, small-world, and hierarchically modular [26,27,32-34]. Although debate exists [35,36], it is generally believed that these properties may be related to the robustness and stability of the underlying systems [26,27,32-34]. For example, a small-world network has a small diameter and a large clustering coefficient (see Methods), which is believed to be related to an efficient and controlled flow of information [26,34]. In a scale-free network, the probability for a node to have k edges follows a power-law distribution, i.e. $P(k) = c \times k^{-\gamma}$. The implication of the scale-free property is that a few nodes in the network are highly connected, acting as hubs, while most nodes have low degrees. Scale-free networks are believed to be robust to random failures, but vulnerable to deliberate attacks [27,32]. In comparison, in a random network (specifically, an Erdos-Renyi random network [26]), connections are spread almost uniformly across all nodes [26,34]. Furthermore, although a random network may also have a small diameter, it usually has a near zero clustering coefficient [26,34]. Several studies have analyzed the value-based gene co-expression networks, and reported some interesting but controversial results [11-13,25]. Here we analyze the

topologies of both the rank-based and the value-based networks, and compare with previous results.

We obtained a set of yeast gene expression data measured in 173 different time points under various stress conditions [37], and selected 3000 genes that showed the highest variations. We constructed four gene co-expression networks using the rank-based method with $d = 2, 3, 4$ and 5 , respectively. For each rank-based network, we constructed two random networks as follows. First, we randomly permuted the expression data of each gene independently, and constructed a rank-based network using the permuted data. Second, we randomly rewired the connections in a true rank-based network, but preserved the degree for every node [34]. For comparison, we also constructed four value-based networks, using the Pearson correlation coefficient as a similarity measure. The thresholds were chosen such that the average degrees are 10, 30, 50, and 100, respectively, in the resulting networks. Similar to the rank-based networks, we obtained two random networks for each true value-based network, one constructed from randomly permuted data and the other by randomly rewiring the true network.

Table 1 lists some statistics of these networks. In the rank-based networks, almost all genes are linked to the largest component with d as small as 2. Furthermore, compared to both the randomly rewired networks and

the networks constructed from randomly permuted data, the true rank-based co-expression networks have slightly larger average path lengths and diameters, but much larger clustering coefficients, indicating that the rank-based co-expression networks have the small-world property. In contrast, the true value-based co-expression networks contain many singletons. For example, with a Pearson correlation coefficient threshold of 0.69, about 900 genes are singletons, even though the average node degree is much higher than in the rank-based networks. Furthermore, although the value-based networks have high clustering coefficients, their randomly rewired counterparts have almost similarly high clustering coefficients. This observation suggests that the high clustering coefficient of the value-based networks is partially because their non-singleton nodes are almost completely connected, in which case the structure cannot be destroyed by any random rewiring.

Figure 3(a) and 3(b) shows the degree distributions of these networks. As indicated by a linear relationship in the log-log plot, the rank-based networks constructed from the real data exhibit a power-law degree distribution for all the d values considered. This suggests that an overall scale-free topology is a fairly robust feature of the co-expression networks. In contrast, the networks constructed from randomly permuted gene expression data contain significantly fewer high-degree nodes, and exhibit exponential degree distributions. The value-based networks appear to follow power-law degree distributions as well; however, they have a much larger

number of high degree nodes than the rank-based networks.

To quantify the difference between the degree distributions of the value-based and rank-based networks, we fitted a power-law function for the degree distribution of each network to determine its γ parameter. The values of γ in the rank-based networks are consistently between two and three. This is typical in many biological networks such as PPI networks and metabolic networks, as well as in real-world social and technology networks [26,34]. In comparison, the γ values in the value-based networks are below one (Figure 3b). Theoretically, it is known that a scale-free network with $\gamma < 2$ has no finite mean degree when its size grows to infinity, and is dominated by nodes with large degrees [26]. Therefore, small values of γ for co-expression networks were reported in several previous studies as a significant difference between the co-expression networks and other biological networks [15,17]. Our results suggest that this difference may simply be an artifact of the network construction method. Consider that genes in some modules are strongly co-expressed with one another, while genes in some other modules are weakly co-expressed. Using the value-based method, when the similarity cutoff is gradually decreased, the genes within the strongly co-expressed modules will be first connected, up to a point that they are almost completely connected, before any gene in the weakly co-expressed modules can be connected to their within-module partners. As a result, the co-expression network will have

Table 1 Statistics of yeast co-expression networks

| | Rank-based networks | | | | | | | | | | | |
|---------------------------|------------------------|--------|---------|----------|----------------------------|------|------|------|--------------------------|-------|-------|-------|
| | Network from real data | | | | Network from permuted data | | | | Randomly rewired network | | | |
| d | 2 | 3 | 4 | 5 | 2 | 3 | 4 | 5 | - | - | - | - |
| Number of singletons | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 | 0 |
| Size of largest component | 2971 | 3000 | 3000 | 3000 | 3000 | 3000 | 3000 | 3000 | 3000 | 3000 | 3000 | 3000 |
| Average degree | 3.6 | 5.4 | 7.2 | 9.0 | 2.8 | 4.0 | 5.2 | 6.3 | 3.6 | 5.4 | 7.2 | 9.0 |
| Largest degree | 22 | 35 | 49 | 57 | 10 | 13 | 14 | 17 | 22 | 35 | 49 | 57 |
| Clustering coefficient | 0.17 | 0.19 | 0.21 | 0.22 | 0.01 | 0.02 | 0.02 | 0.02 | 0.001 | 0.002 | 0.004 | 0.005 |
| Average path length | 8.8 | 6.8 | 5.9 | 5.4 | 9.6 | 6.4 | 5.3 | 4.7 | 6.0 | 4.6 | 4.1 | 3.7 |
| Diameter | 19 | 19 | 14 | 11 | 17 | 10 | 8 | 7 | 12 | 8 | 7 | 6 |
| | Value-based networks | | | | | | | | | | | |
| | Network from real data | | | | Network from permuted data | | | | Randomly rewired network | | | |
| Correlation cutoff | 0.85 | 0.79 | 0.76 | 0.69 | 0.21 | 0.18 | 0.16 | 0.14 | - | - | - | - |
| Number of singletons | 2163 | 1586 | 1311 | 893 | 0 | 0 | 0 | 0 | 2163 | 1586 | 1311 | 893 |
| Size of largest component | 486 | 756 | 1609 | 2055 | 3000 | 3000 | 3000 | 3000 | 835 | 1412 | 1689 | 2107 |
| Average degree | 10(37) | 30(64) | 50(107) | 100(142) | 10 | 30 | 50 | 100 | 10 | 30 | 50 | 100 |
| Largest degree | 211 | 340 | 418 | 581 | 24 | 55 | 79 | 136 | 211 | 340 | 418 | 581 |
| Clustering coefficient | 0.52 | 0.59 | 0.62 | 0.64 | 0.03 | 0.04 | 0.05 | 0.07 | 0.31 | 0.34 | 0.34 | 0.34 |
| Average path length | 2.6 | 2.6 | 6.0 | 4.8 | 3.8 | 2.7 | 2.4 | 2.0 | 2.5 | 2.4 | 2.3 | 2.2 |
| Diameter | 9 | 9 | 18 | 13 | 6 | 4 | 3 | 3 | 5 | 5 | 5 | 5 |

Numbers in parentheses are for non-singleton nodes only.

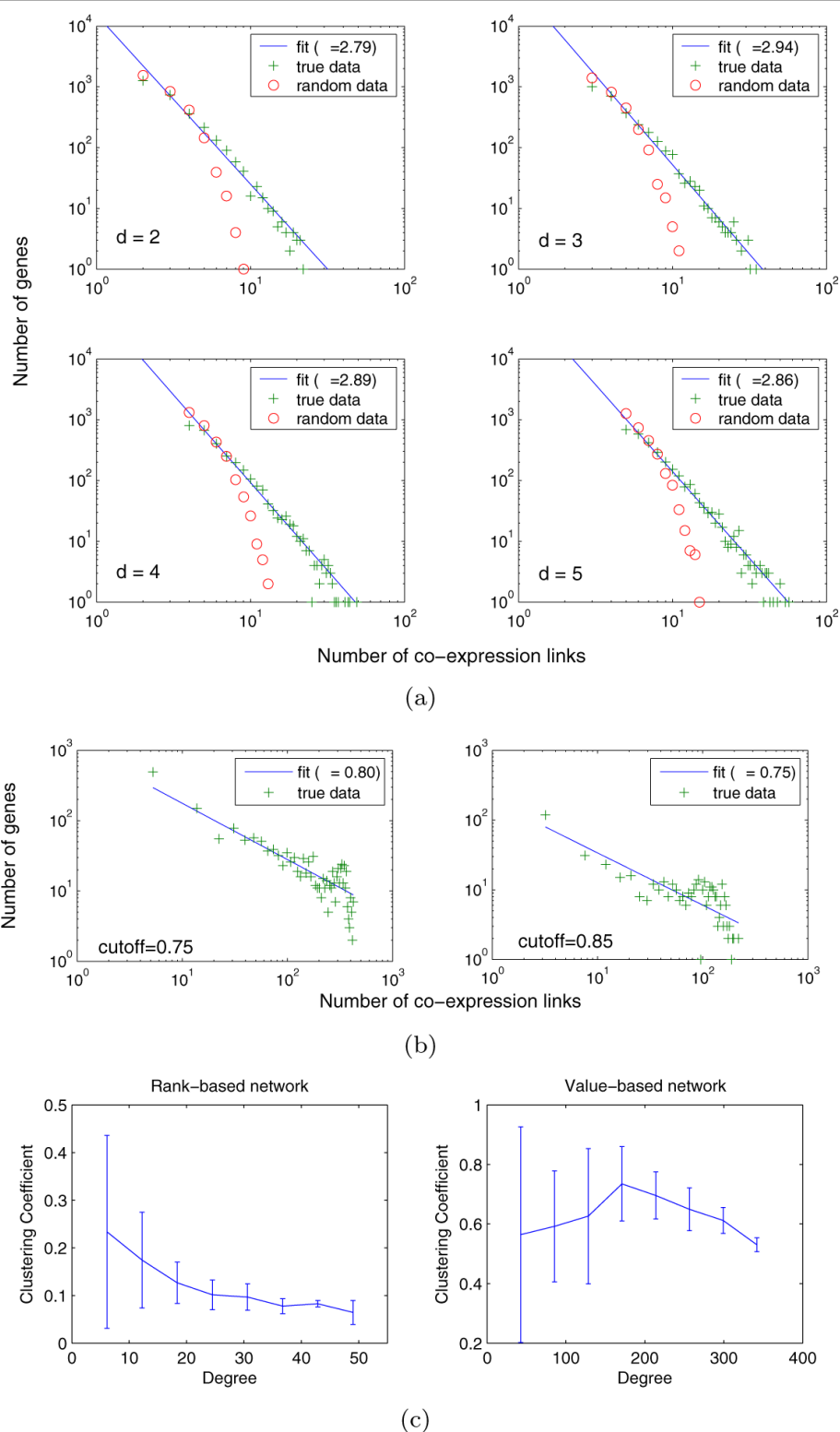


Figure 3 Topological properties of co-expression networks. (a) Degree distribution of rank-based co-expression networks. (b) Degree distribution of value-based co-expression networks. (c) Relationship between clustering coefficient and degree in rank-based and value-based co-expression networks.

many genes with large degrees, resulting in a small slope in the log-log plot. In contrast, with the rank-based method, genes in both strongly and weakly co-expressed modules can be connected, as essentially a different similarity threshold is used for each gene. Therefore, rank-based networks can usually capture the topology of both strongly and weakly co-expressed modules, while value-based networks are often dominated by the strongly co-expressed modules.

Moreover, previous studies have reported that gene co-expression networks lack the hierarchically modular property [11,12]. This property is characterized by a reciprocal relationship between a node's degree and its clustering coefficient [33]. Again, we have found that this claim only applies to the value-based networks. As shown in Figure 3(c), there is a clear reciprocal relationship between the node degree and node clustering coefficient in the rank-based networks, when compared to the value-based networks. This suggests that gene co-expression network can also have hierarchical structures.

Together, these experiments show that the rank-based co-expression networks have all the common topological properties of many other biological networks, while the value-based networks seem to differ significantly. Although these do not necessarily prove that rank-based networks are biologically more meaningful than value-based networks, the former seems to be able to capture the underlying topological structures better.

Module discovery and analysis in gene co-expression networks

Gene co-expression networks with thousands of nodes are difficult to visualize and comprehend. A useful strategy for analyzing such a network is to partition it into subnetworks, where the nodes within each subnetwork are relatively densely connected to one another but have fewer connections to the other subnetworks. In gene co-expression networks, such subnetworks can be considered as candidates of functional modules, as genes within each subnetwork are mutually co-expressed, while co-expression between genes in different subnetworks are sparse. Many graph partitioning algorithms have been developed in computer science [38]. Similar to clustering, one major difficulty in graph partitioning is to determine the number of partitions. Some methods do not require this to be explicitly determined in advance, but require other parameters, which are also difficult to obtain. For example, MCL, one of the best graph partitioning algorithms, requires an inflation parameter, and setting the parameter to different values may result in very different results [29].

To address this difficulty, we introduce an algorithm that we have developed recently for identifying "communities" in arbitrary networks [28]. The main motivation for the algorithm is that each "community", or a

subnetwork, must contain more intra-community edges than would be expected by chance if the connections were random. With this motivation, we developed an algorithm to optimize an objective function called modularity, which is precisely defined as the percentage of intra-community edges minus the random expectation (see Methods). The algorithm, named *Qcut*, has been shown to be effective in finding statistically significant and practically interesting graph partitions in many synthetic networks, social networks, and biological networks, without any user-tunable parameters, and has outperformed the existing algorithms based on similar motivations [28].

We evaluate the performance of *Qcut* on gene co-expression networks in several ways. We first use synthetic microarray data where the true modular structure is known, so that we can directly measure the accuracy. We then use two real microarray data sets to evaluate the overall biological significance of the identified gene modules, with two different metrics. The first metric is a commonly used approach based on the enrichment of specific Gene Ontology terms in the modules, which may be biased by the number of modules and module sizes. The second is a new metric that we introduced based on the idea of reference networks, which can be obtained from a variety of sources, such as gene annotations or protein-protein interaction networks (See methods).

Evaluation using synthetic microarray data

To objectively evaluate the accuracy of the modules detected by *Qcut*, we tested it on a large collection of synthetic gene expression data. The data sets, available at <http://www.biostat.pitt.edu/bioinfo/publication.htm>, were used to evaluate many clustering algorithms in a previous study [39]. Each data set contains simulated expression data of approximately 600 genes under 50 conditions. Each gene was pre-assigned to one of fifteen clusters, and the genes in the same cluster had their expression profiles generated from a common log normal distribution. Gaussian noises were then added to the data set to simulate experimental noises. A higher level of Gaussian noise generally makes the data more difficult to cluster. Since the correct clusters are known, we used a well-known metric called the adjusted Rand Index to measure the accuracy of *Qcut* (see Methods) [40].

We first compared the accuracy of *Qcut* on co-expression networks constructed by three methods: value-based, rank-based, and CLR [19]. We used Euclidean distance as the basis to measure the dissimilarity between two genes. For the value-based method, we normalized the distance to be between 0 and 1, and constructed a series of co-expression networks for each data set using different threshold values. As shown in

Figure 4(a), the threshold that results in the best clustering accuracy varies for different data set. For more noisy data set a larger threshold value is needed, which suggests that choosing a right threshold is critical for the value-based method. The CLR method, in contrast, by converting the raw distances to z-scores, effectively removed such dependency and the best clustering accuracy is achieved at the same z-score equal to 2, corresponding to a p-value 0.05, for all data sets (Figure 4b). Interestingly, for the rank-based method, the clustering accuracy is almost invariant for rank cutoffs between 2 and 8 (Figure 4c). Figure 4(d) shows the best accuracy that can be achieved on the three types of networks. As can be seen, the rank-based networks clearly have the highest accuracy for intermediate levels of noises ($SD = 0.4$ or 0.8). For data with lower noises, all three methods resulted in perfect accuracy, and for data set with the highest level of noise ($SD = 1.2$), all three methods converges to about the same accuracy. Next we compared the clustering accuracy of *Qcut* on rank-based networks with several widely-used clustering algorithms including *k*-means clustering, hierarchical clustering [1], and tight clustering [39], applied directly to the gene expression data without deriving co-expression networks. In this test, *Qcut* was applied to rank-based co-expression networks constructed using d values equal to 4. In addition, we also tested one of the best graph partitioning algorithms called the Markov Clustering algorithm (MCL) [29], which is applied to rank-based networks as well. Since the results of MCL depend heavily on the choice of an inflation parameter (I), we applied MCL to the rank-based networks constructed with d fixed at 4, but varied I from 1.3 to 1.7, with an increment of 0.1, and took the best clustering accuracy resulted from these parameters. We used the MATLAB (the MathWorks Inc.) implementation of the *k*-means and hierarchical clustering algorithms. *k*-means clustering was run with k equal to 15, and was repeated 50 times for each data set to obtain the best results. The hierarchical clustering was performed using average linkage, and the final cluster tree was cut at an appropriate depth to generate 15 clusters. The accuracies of tight clustering were directly obtained from the original study that was done on the same data set [39]. Our evaluation results show that, even without any parameter tuning, *Qcut* outperformed the competing algorithms in identifying the true modular structures embedded within the synthetic microarray data. As shown in Figure 5, the clustering accuracy of *Qcut* is clearly better than that of the hierarchical clustering and tight clustering. The accuracy of *Qcut* is similar to *k*-means, except for the data sets of the highest level of noise. The synthetic data set with the highest level of noise may represent an extreme case in practice, as many of the clusters in this data set are not

distinguishable visually (Figure S1 in Additional File 1). However, *k*-means achieved this accuracy with the number of clusters given explicitly, while *Qcut* did not have this information at all. In these synthetic data sets, the number of clusters is the single most important parameter and *k*-means is expected to work well when that is known. We tried to combine *k*-means with several popular methods to automatically determine the number of clusters, including the gap statistic [41], Silhouette [42], and the Dunn's Index [43]. Our results suggest that if the values of k are automatically determined, *k*-means performs much poorer than our method, especially for data sets with $SD \geq 0.4$ (Figure 5). The results of MCL are two-fold. On one hand, when an appropriate inflation parameter is chosen ($I = 1.5$ in this experiment), MCL has an accuracy similar to that of *Qcut*, except for the data set with the highest noises, indicating a superior performance of graph-based algorithms in general. On the other hand, the accuracy of MCL depends on the choice of the inflation parameter, and may be much lower than that of clustering algorithms if a suboptimal inflation parameter is used (data not shown).

Functional modules in a yeast gene co-expression network

To evaluate the performance of our algorithm on real biological data, we applied *Qcut* to cluster the four rank-based yeast co-expression networks constructed in the previous subsection. The best numbers of clusters suggested by *Qcut* for the four networks are 24, 20, 12 and 12, respectively. As shown on synthetic data, when the number of clusters is known, the clustering algorithms that explicitly use this information, such as *k*-means, perform better than those without this information, such as the hierarchical clustering. Therefore, here we compared the performance of *Qcut* to *k*-means and two other popular clustering algorithms, namely self-organizing maps (SOM) [44] and spectral clustering [45], both of which require the number of clusters to be given. Applying the value-based method to the yeast data set, as we showed in Figure 1, the network either contains too many singletons, or are too densely connected to be partitioned by *Qcut* (or any other graph algorithms). Therefore, we did not evaluate its accuracy on the real data set. Interestingly CLR had similar behavior as the value-based method on the yeast data set, indicating that it is partially value-based.

We used the SOM implementation in the microarray software suite TM4 from TIGR, and implemented our own version of spectral clustering. *K*-means, SOM and spectral clustering were applied directly to the expression data, using Pearson correlation coefficient as the similarity metric. We obtained 24, 20, 12 and 9 clusters using each of the three competing algorithms. To test if the competing algorithms may give a better result with

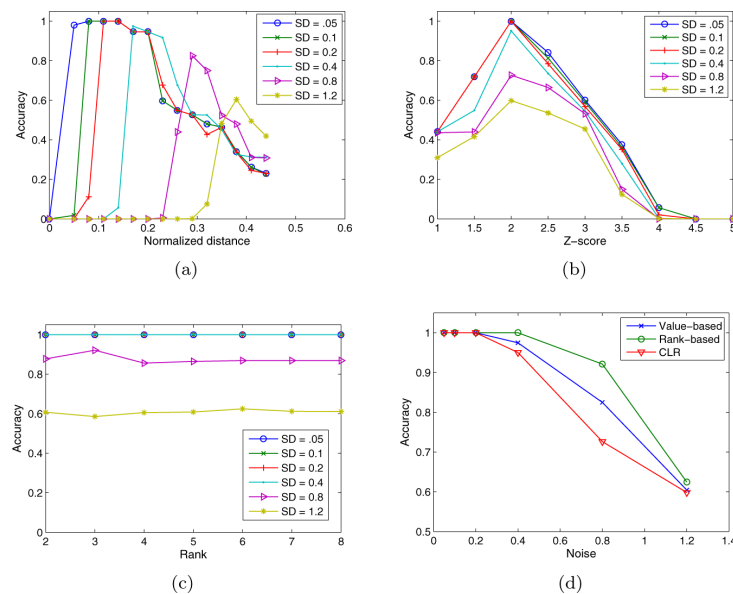


Figure 4 Effects of network construction methods on the clustering accuracy of Qcut. (a) Clustering accuracy on value-based networks, as a function of the distance cutoffs. (b) Clustering accuracy on CLR co-expression networks, as a function of the Z-score cutoffs. (c) Clustering accuracy on rank-based networks, as a function of the rank cutoffs. (d) Best clustering accuracy on the three types of networks, constructed with the optimal cutoffs. In all four plots, each data point is an average over the results of 100 synthetic microarray data sets.

a different setting of the number of clusters, we also applied the spectral clustering with the number of clusters k equal to 5, 6, ..., 25. SOM was executed on 4×6 , 4×5 , 3×4 , and 3×3 grids to produce the desired number of clusters. Because *Qcut* identified 12 clusters on both the $d = 4$ and $d = 5$ networks, we compared the 12 clusters identified from the $d = 5$ network by *Qcut* with the 9 clusters identified by the competing algorithms to avoid redundant comparison. Another reason for this comparison is that *Qcut* often produces a few small clusters, while the clusters of the competing algorithms are relatively uniform in size. Therefore, the “effective” number of clusters is smaller for *Qcut* than for other algorithms, so we used this comparison to compensate for the differences in cluster sizes.

To validate the biological significance of the clusters, we first counted the number of Gene Ontology (GO) terms enriched in the clusters and the percentage of clusters that had at least one enriched GO term, at various significance levels. As shown in Figure 6, the clusters identified by *Qcut* contain more enriched GO terms than the competing algorithms for most of the p -value cutoffs (Figure 6(a)-(d)). Furthermore, the percentage of clusters containing at least one enriched GO term is also higher for *Qcut* than for the other algorithms (Figure 6(e)-(h)).

Second, as the above measurement may be biased by the number of clusters or the cluster size distributions, we compared these algorithms using a novel metric

based on three reference networks that capture different functional interactions between genes: a co-annotation network based on GO terms of biological processes, a co-regulation network based on ChIP-chip data, and a PPI network (see Methods). While the PPI network has unweighted edges, the GO-based and ChIP-based reference networks have weighted edges. We discretized the GO-based and ChIP-based reference networks using a series of weight cutoffs. We then scored the gene modules identified by each algorithm using these reference networks (see Methods). As shown in Figure 7 and Figure S2 in Additional File 1, the gene modules identified by *Qcut* always had the highest score, using all three types of reference networks. The spectral clustering algorithm performed better than the other clustering algorithms. As *Qcut* and the spectral clustering algorithm share some similar spirit in capturing the topology embedded in a data matrix, this result seems to suggest that topological features are important for achieving good clustering results on gene expression data. We also randomly shuffled the gene modules identified by *Qcut* while preserving the sizes of the modules, and scored the random clusters using the reference networks. The scores for the random modules are very close to zero in all cases (Figure 7 and Figure S2 in Additional File 1), indicating that the results obtained by

Qcut and the clustering algorithms are not due to chance
The modules in the $d = 2$ co-expression network have the worst scores according to all three reference

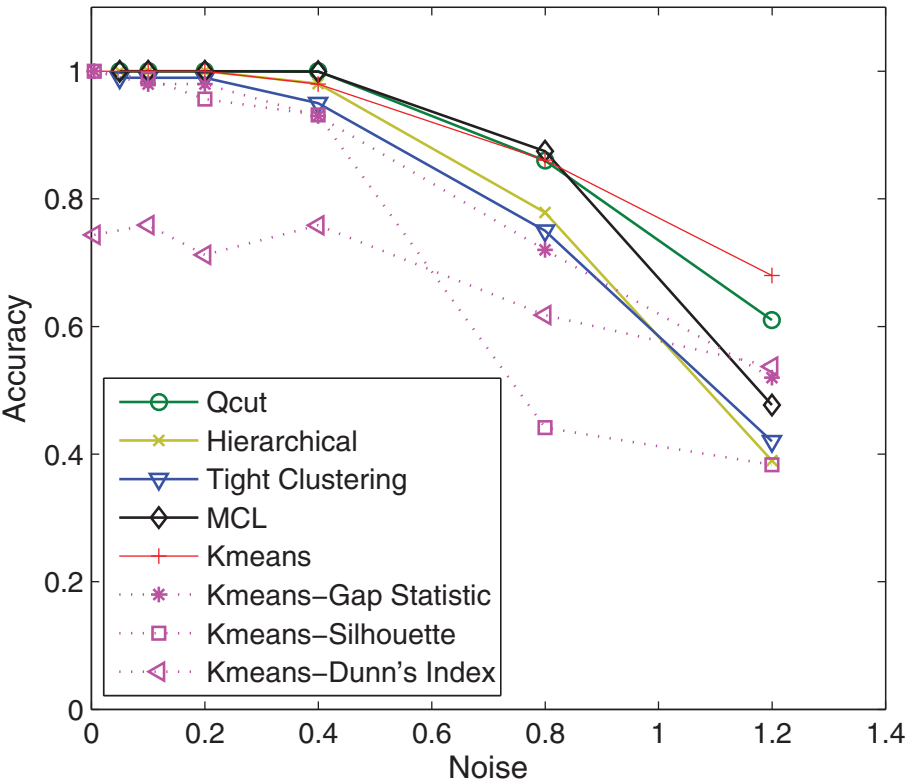


Figure 5 Comparison of different clustering methods using synthetic microarray data. *Qcut* and *MCL* are applied to rank-based networks constructed with $d = 4$. Each data point in the plot is an average over 100 synthetic microarray data sets.

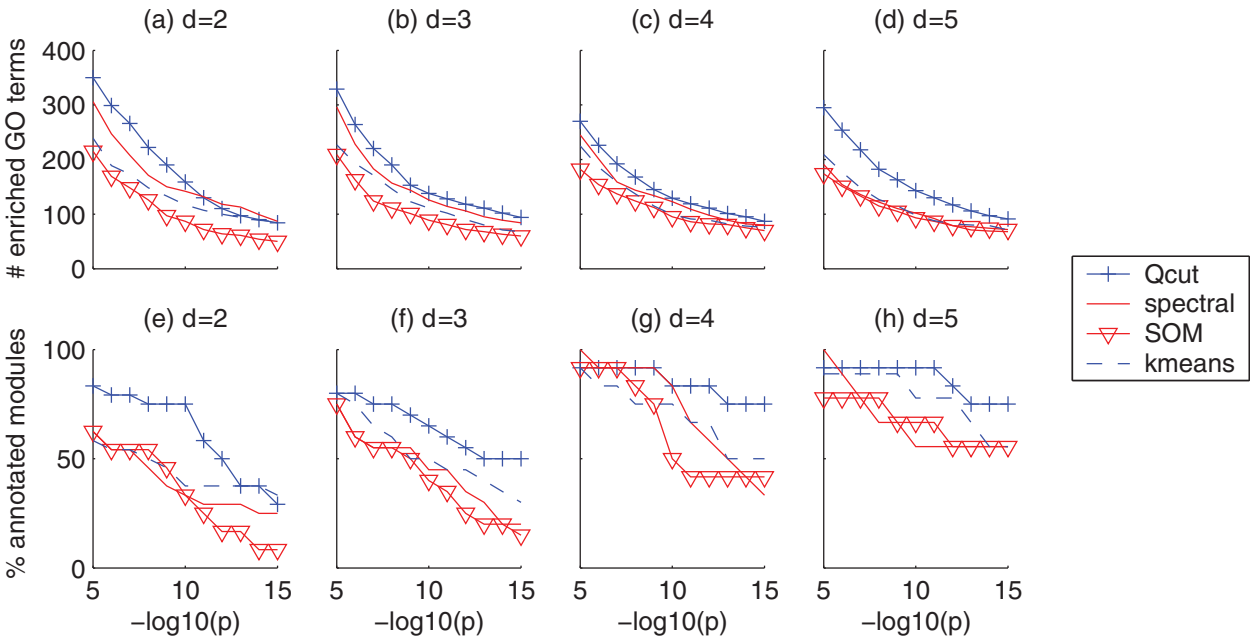


Figure 6 Enrichment of GO terms in yeast co-expression networks. Vertical axes in (a)-(d): number of GO terms enriched in the clusters. Vertical axes in (e)-(h): percentage of clusters that are enriched with at least one GO term. Horizontal axes: p -value cutoff to consider a GO term enriched.

networks (Figure 7 and Figure S2 in Additional File 1), indicating that the $d = 2$ network might be too sparse to capture all functional relationships. The $d = 4$ co-expression network has the highest scores according to all three reference networks, while the networks with $d = 3$ or 5 give slightly worse results. As stated above, to test if the competing algorithms may give a better result with a different setting of the number of clusters, we applied the spectral clustering, which has a better accuracy than the other two algorithms, with the number of clusters k equal to 5, 6, ..., 25. We scored the gene modules using the GO-based reference network at a weight cutoff of 0.8. As shown in Figure S3 in Additional File 1, the best module score achieved by spectral clustering is 0.323 (at $k = 13$), which is significantly lower than the module score by *Qcut* ($Q = 0.384$). This shows that our evaluation results have not been biased by the number of clusters.

In addition, we combined k -means with gap-statistic [41], which estimated the optimal number of clusters to be 6. We also tested SOTA [46], a hierarchical version of SOM that can automatically determine the number of clusters. SOTA returned 11 clusters, a number similar to our optimal number. However, only 6 of the 11 clusters had significantly enriched functions, and the most significant function had a p -value 10^{-49} , as compared to 10^{-106} in our method (Table 2 and Table S1 in Additional File 1).

Table 2 summarizes the modules identified by *Qcut* from the $d = 4$ network. For each module, we show the

most significantly enriched GO biological process terms and the transcription factors whose targets are significantly enriched in the module (see Methods). As shown, most modules contain highly coherent functional terms, and are co-regulated by common transcription factors. For example, the majority of genes in module 12 are involved in protein biosynthesis ($p = 10^{-85}$), and can be bound by FHL1 ($p = 10^{-105}$) and RAP1 ($p = 10^{-48}$), both of which are known to be involved in rRNA processing and regulating ribosomal proteins [47]. Module 9 is significantly enriched with genes that are involved in generation of precursor metabolites and energy ($p = 10^{-33}$), and can be bound by HAP4 ($p = 10^{-16}$), a TF regulating carbohydrate metabolism [47]. Module 2 contains almost two-thirds of the ribosome biogenesis genes ($p = 10^{-106}$), but no TFs were found to bind to this set of genes specifically. Module 11 is enriched with genes that can be bound by eight different TFs. Interestingly, all eight TFs are known cell-cycle regulators [47]. Several small modules correspond to specific functional groups. For example, 17 of the 22 genes in module 10 are involved in Ty element transposition ($p = 10^{-29}$). Half of the 18 genes in module 3 are related to chromatin assembly or disassembly ($p = 10^{-13}$); six of them are regulated by transcription factors HIR1/2/3, which are known to be involved in the transcription of histone genes [47]. Modules 5 and 7 contain both a large fraction of genes with unknown functions and groups of genes with significantly enriched common functions or common TFs. It is possible that these uncharacterized

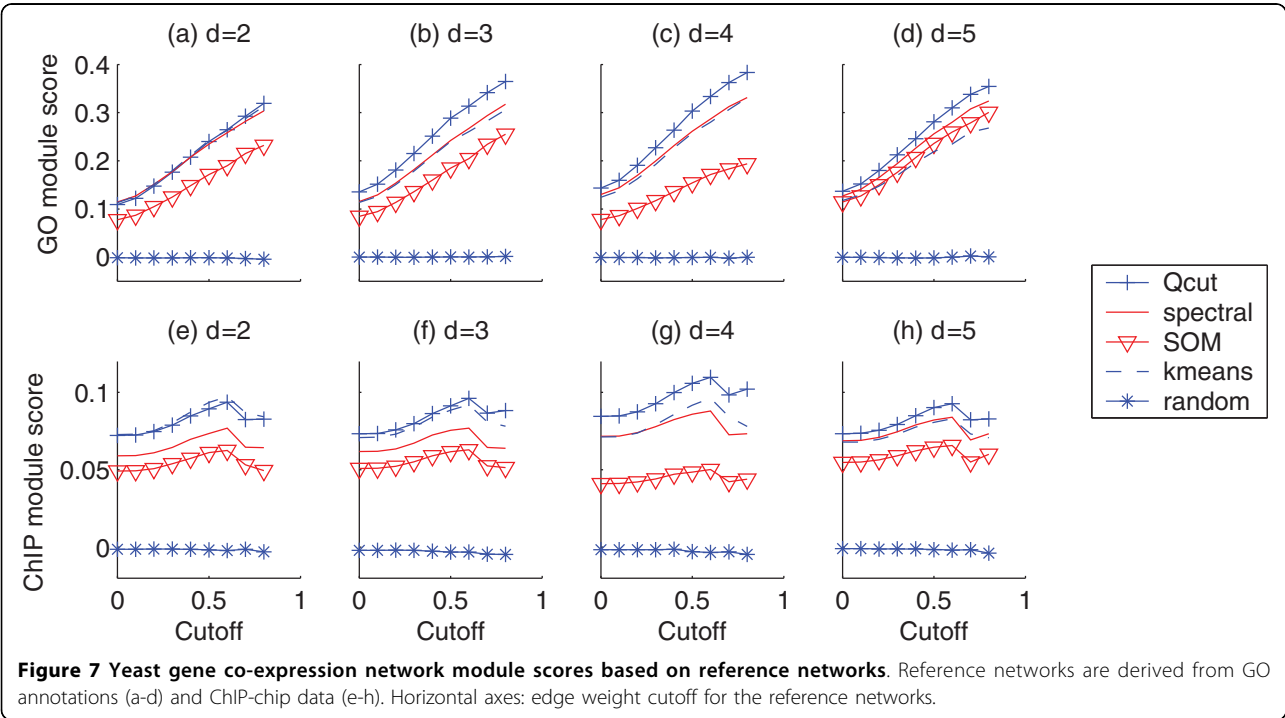


Table 2 Functional modules in a yeast co-expression network

| Cluster | Size | Category ¹ | Term | Count | Enrichment ² | P-value |
|---------|------|-----------------------|--|-------|-------------------------|----------|
| 1 | 361 | BP | protein catabolism | 32 | 4.2 | 2.0E-12 |
| | | BP | protein folding | 21 | 5.9 | 1.6E-11 |
| 2 | 498 | BP | ribosome biogenesis | 133 | 9.2 | 2.0E-106 |
| 3 | 18 | BP | chromatin assembly or disassembly | 9 | 36.4 | 5.3E-13 |
| | | TF | HIR2 | 6 | 129.8 | 2.3E-12 |
| | | TF | HIR1 | 6 | 62.9 | 3.0E-10 |
| | | TF | HIR3 | 6 | 57.7 | 5.3E-10 |
| 4 | 25 | BP | telomerase-independent telomere maintenance | 4 | 82.3 | 1.1E-07 |
| | | BP | biological process unknown | 16 | 2.9 | 7.6E-06 |
| | | TF | GAT3 | 13 | 56.8 | 3.5E-21 |
| | | TF | YAP5 | 15 | 43.5 | 5.8E-17 |
| | | TF | PDR1 | 9 | 25.8 | 3.1E-11 |
| | | TF | MSN4 | 8 | 35.0 | 3.8E-11 |
| 5 | 422 | BP | spore wall assembly | 16 | 7.0 | 1.6E-10 |
| | | BP | biological process unknown | 138 | 1.5 | 1.2E-07 |
| | | TF | NRG1 | 21 | 4.2 | 1.4E-08 |
| | | TF | SUM1 | 16 | 3.9 | 2.3E-06 |
| | | TF | PHD1 | 15 | 3.4 | 3.2E-05 |
| 6 | 99 | - | - | - | - | - |
| 7 | 463 | BP | carbohydrate metabolism | 41 | 2.9 | 4.6E-10 |
| | | BP | biological process unknown | 178 | 1.7 | 9.5E-17 |
| | | BP | response to stimulus | 62 | 1.7 | 2.0E-05 |
| | | TF | UME6 | 25 | 2.5 | 2.6E-05 |
| | | TF | NRG1 | 15 | 2.8 | 3.6E-04 |
| 8 | 108 | BP | nitrogen compound metabolism | 25 | 7.0 | 5.2E-15 |
| | | TF | MET31 | 4 | 9.6 | 8.0E-04 |
| | | TF | MET32 | 5 | 5.7 | 2.1E-03 |
| 9 | 192 | BP | generation of precursor metabolites and energy | 50 | 8.2 | 7.5E-33 |
| | | TF | HAP4 | 22 | 9.2 | 5.1E-16 |
| 10 | 22 | BP | Ty element transposition | 17 | 58.6 | 6.2E-29 |
| | | TF | SUM1 | 4 | 18.9 | 5.8E-05 |
| 11 | 604 | BP | carboxylic acid metabolism | 76 | 3.0 | 2.4E-19 |
| | | BP | cell organization and biogenesis | 212 | 1.6 | 3.7E-15 |
| | | TF | SWI6 | 45 | 2.9 | 3.6E-11 |
| | | TF | SWI4 | 44 | 2.8 | 2.7E-10 |
| | | TF | FKH2 | 35 | 3.0 | 4.7E-09 |
| | | TF | MBP1 | 36 | 2.8 | 1.9E-08 |
| | | TF | STE12 | 22 | 3.6 | 7.9E-08 |
| | | TF | NDD1 | 30 | 2.9 | 1.1E-07 |
| | | TF | FKH1 | 34 | 2.5 | 9.6E-07 |
| | | TF | MCM1 | 22 | 2.9 | 3.9E-06 |
| 12 | 186 | BP | protein biosynthesis | 131 | 6.4 | 6.4E-85 |
| | | TF | FHL1 | 96 | 17.1 | 3.3E-105 |
| | | TF | RAP1 | 58 | 11.5 | 2.2E-48 |

¹ For each cluster, significantly enriched biological process GO terms (BP) or binding of transcription factors (TF) are counted.

² Fold of enrichment is calculated as: $\frac{(\text{number of genes in cluster with the term}) \times (\text{number of genes in genome})}{(\text{number of genes in cluster}) \times (\text{number of genes in genome with the term})}$

genes also have the functions that are enriched in the module.

We also found a very interesting small module that may deserve further investigation. Among the 25 genes in module 4, four genes have a common function in telomere maintenance ($p = 10^{-7}$). While the majority (16) of the remaining genes encode hypothetical proteins and have unknown functions ($p = 10^{-6}$), a careful inspection showed that all four characterized genes and five of the 16 uncharacterized ones are located near telomeric regions [47]. Moreover, a significant number of genes in this module are regulated by four common transcription factors: GAT3 ($p = 10^{-21}$), YAP5 ($p = 10^{-17}$), PDR1 ($p = 10^{-11}$), and MSN4 ($p = 10^{-11}$) (Figure 8 and Table 2). Our results suggest that these uncharacterized genes as well as the four transcription factors are very likely to be involved in the function or maintenance of telomere, which has not been reported in the literature.

Functional modules in an Arabidopsis gene co-expression network

To test our method on high organisms, we applied it to a set of Arabidopsis gene expression data downloaded from the AtGenExpress database <http://www.uni-tuebingen.de/plantphys/AFGN/atgenex.htm>. We studied the cold stress response, for which the dataset contains the expression of 22 k Arabidopsis genes in the root and shoot tissues in six time points following cold stress treatment and under the normal condition. We selected 2545 genes that are up- or down-regulated by at least five-folds in at least one of the six time points in either tissue. We then constructed a co-expression network by connecting each gene to its top three correlated genes (i.e. $d = 3$), resulting in 5838 co-expression links in total.

Our clustering algorithm partitioned the network into 19 modules, with a Q value of 0.81, indicating a strong modular structure. Similar to the previous experiment, we first examined the GO terms enriched in the clusters

at various significance levels, and compared them with the results of the k -means algorithm with $k = 19$. As shown in Figure 9(a) and 9(b), the modules identified by our method contain significantly more enriched GO terms than that identified by k -means, and GO terms are enriched in more modules in our method than in k -means. Furthermore, our method achieved significantly higher module scores in the GO-based reference networks than the k -means algorithm (Fig 9(c)). Table S2 in Additional File 1 lists the most enriched functional categories for each module. Several modules are enriched with functions that are known to be related to cold stress responses, e.g. modules 7 (photosynthesis, $p = 10^{-16}$), 11 (circadian rhythm, $p = 10^{-5}$), 14 (response to heat, $p = 10^{-15}$), 15 (antiporter activity, $p = 10^{-6}$) and 18 (lipid binding, $p = 10^{-8}$). Due to the scarcity of functional annotations in Arabidopsis, the enrichment p -values as well as the module scores of the gene modules are less significant in Arabidopsis than in yeast, which is to be expected.

Cancer subtype classification from sample co-expression networks

In this subsection, we show that our co-expression network analysis method can also be applied to identify sample modules. Conceptually, there is no difference in constructing and analyzing gene or sample co-expression networks: in the latter we treat each sample as a network node, and connect two samples when their expression profiles are similar according to some similarity measure. In practice, however, sample co-expression networks are usually smaller than gene co-expression networks, but the edges in sample co-expression networks may be computed from very high-dimensional data, since each sample is described by thousands of genes. It is therefore interesting to see whether the same network construction and module detection methods can be applied here without much tuning of the

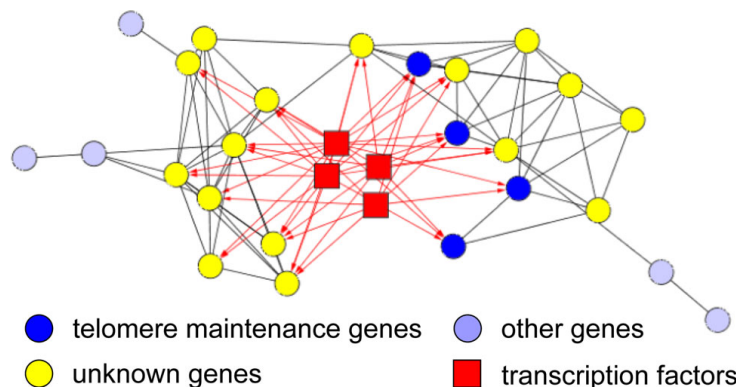
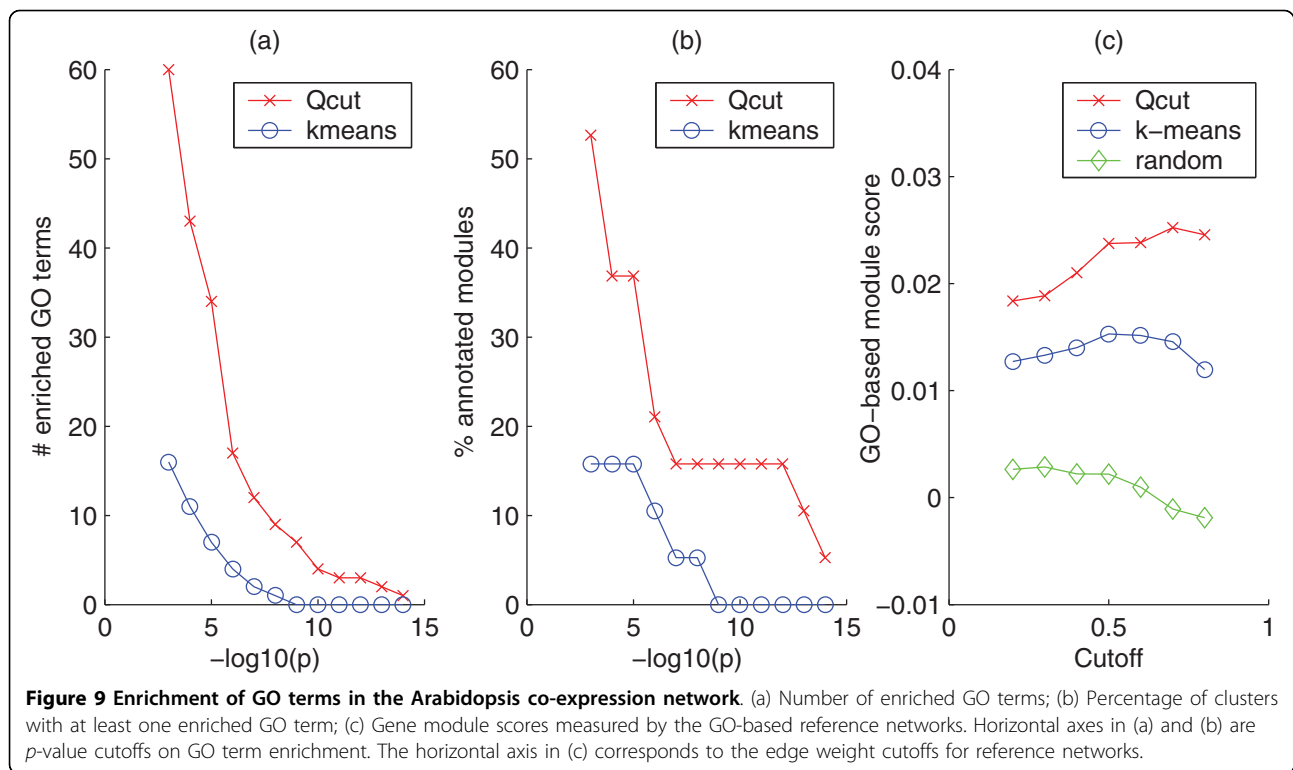


Figure 8 A network of co-expressed and co-regulated genes with functions in telomerase maintenance. Each directed edge pointing from a TF to a gene represents a protein-DNA interaction. All other edges represent co-expression relationships.



parameter, i.e. the value of d . As an application of our approach, we applied our method towards an automatic classification of tumor cells.

An accurate classification of tumor cells is crucial for effective therapy [48]. Traditionally, tumors have been classified by their morphologic appearance, which is, unfortunately, often very subjective. Furthermore, tumors with similar histological features may respond very differently to chemotherapy [30]. To address this problem, a promising alternative or complementary strategy is to classify tumors based on their genetic profiles, i.e. the activity of hundreds or thousands of genes that are involved in the disease. Most of the existing tumor classification approaches are based on supervised learning, such as support vector machines or decision trees, which aimed at identifying genetic features to distinguish two or more known tumor (sub-)types [49,50].

Here, we ask whether it is possible to automatically classify tumor samples in the absence of training data that provide known tumor (sub-)types or even the number of distinct (sub-)types. This unsupervised learning approach has a few advantages over supervised learning methods. First, the existing tumor classifications are based on histological features, which may be unreliable themselves. Second, using unsupervised learning, we may be able to discover novel tumor sub-types that have not been characterized by histological features previously. On the other hand, it is crucial to confirm

whether the automatically discovered tumor (sub-)types are biologically meaningful and indeed provide useful information for understanding the disease mechanism or for improving its treatment.

In this study, we chose to focus on lymphoma, a family of tumors involving cells of the immune system. We obtained a data set containing the expression data of 4026 genes for 96 samples belonging to nine cell types, including three different types of tumors, namely, diffuse large B cell lymphoma (DLBCB), chronic lymphocytic leukemia (CLL), and follicular lymphoma (FL), as well as normal B and T cells at different stages of cell differentiation [30]. We constructed a rank-based network of the samples using Pearson correlation coefficient as the similarity measure and the value of d at 5. Edges with a Pearson correlation coefficient < 0.2 were removed to reduce false connections.

Applying *Qcut* to the network, we identified eight modules. Remarkably, without any prior knowledge about the number of cell types in the samples, our algorithm automatically separated different cell types into different modules, with a few exceptions (Figure 10 and Table S3 in Additional File 1). Furthermore, the results are almost invariant when we varied d between 3 and 7, indicating a very stable modular structure among the samples. As shown in Figure 10, blood T cells and activated blood B cells are perfectly classified into their own modules. CLL and resting B cells are grouped into a

single module, which is not surprising since the CLL has a very low proliferation rate, similar to resting B cells [30]. The two germinal centre B (GCB) cell samples are grouped with the FL cells and are far away from the activated B cells, which confirms the hypotheses that GCB represents a distinct stage of B cell differentiation, and that the FL arises from this stage of B-cell differentiation [30]. The transformed cell line module also contains three DLBCL cells. A closer inspection showed that two of the DLBCB samples in this module (OCI-Ly1 and OCI-Ly3) are laboratory-cultivated cell lines rather than samples from real patients (Figure 10), which may be the reason that they are grouped with the transferred cell lines. The Lymph node/tonsil cells are grouped with DLBCL as in previous studies [30].

Interestingly, the majority of the DLBCB samples are grouped into three modules (DLBCL-1, 2, and 3). It is well known that not all DLBCL tumors are equal: 40% of patients respond well to chemotherapy and have prolonged survival, while the others have a much shorter survival time after treatment [30]. Previous studies have suggested that DLBCL tumors can be categorized into two subtypes: GCB-like DLBCL and activated B-like DLBCL [30]. The GCB-like DLBCL shares gene signatures that distinguished germinal centre B cells from B cells in other stages, while the activated B-like DLBCL shared many gene signatures with normal lymph node and tonsil. On average, GCB-like DLBCL patients have a much higher survival rate than activated B-like DLBCL patients after comparable chemotherapy [30]. In our sample co-expression network, the GCB cells connect to DLBCL-1 cells while the tonsil and lymph node cells fall in the DLBCL-2 modules, suggesting that the two modules may correspond to the two well-known DLBCL subtypes. Indeed, the median survival durations for the patients in the DLBCL-1 and DLBCL-2 modules are 71 and 22 months, respectively. Furthermore, 11 out of 15 (73%) patients in the DLBCL-1 module lived more than five years after treatment, while only 2 out of 13 (15%) patients in the DLBCL-2 module survived that long. Even more interestingly, however, the DLBCL-3 module, which is apart from GCB and Tonsil/Lymph nodes, has the lowest survival rate overall. The median survival durations for this module is 12 months, and only 1 out of 10 (10%) patients in this module survived more than five years after treatment. The lack of GCB-like or activated B-like signatures and the lower survival rate seems to suggest DLBCL-3 to be a separate subtype.

Conclusions

In this paper, we presented a general co-expression network-based approach for the analysis of high-throughput gene expression data. We introduced a simple rank-

based method to construct gene or sample co-expression networks, and an algorithm called *Qcut* to identify modules from the co-expression networks. We also introduced a novel metric to evaluate the functional significance of gene modules based on reference networks.

On synthetic data, we showed that our method can automatically discover the embedded modular structures and that our method significantly outperformed a number of competing algorithms. We applied our method to two real data sets, and showed that it can produce statistically more significant gene functional modules than conventional clustering methods such as *k*-means and self-organizing maps, evaluated by a number of criteria. Furthermore, our test on a sample co-expression network showed excellent results in separating different types or subtypes of tumor cells. All these were achieved without knowing the number of modules in advance.

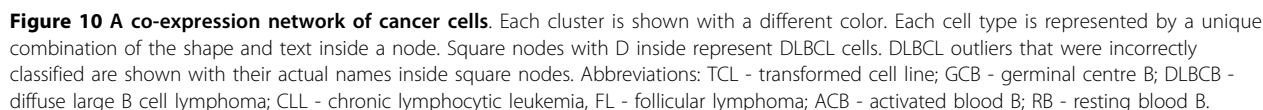
We reported several interesting biological results, including some testable biological hypotheses. We showed that the rank-based co-expression networks have all the common topological properties that exist in other biological networks, challenging the previous results on the topology of gene co-expression networks. We discovered an interesting gene module in yeast suggesting that some uncharacterized genes and transcriptional regulators may be involved in maintaining telomere integrity. We also identified a module of tumor cells which seems to correspond to a new subtype that has not been identified before.

Although we have only demonstrated our method on gene expression data, it can be applied to other types of experimental data as well. The rank-based network construction method can be used to capture the relationships among other entities. The efficiency of our module detection method and its parameter-free feature make it well suited for identifying intrinsic structures in large-scale network data. Another advantage of our method is its flexibility in integrating different types of information. For example, co-expression networks and transcriptional regulatory networks can be easily integrated; the modules in those integrated networks will then represent co-expressed and co-regulated genes, as well as their regulators. Similarly, we would expect other types of information, such as protein-protein interaction, phylogeny and phenotypes, to be integrated to produce more biologically insightful results.

Methods

Network topological analysis

The topological properties of networks were analyzed using MATLAB (the MathWorks Inc.). For each network the number of nodes and number of edges was simply counted. A singleton is a node with zero connection. The average degree $\langle k \rangle$ was calculated as the



Our module detection algorithm, *Qcut*, optimizes a modularity function Q to automatically determine the most appropriate number of modules in a network. The algorithm can handle networks of several thousands of nodes in a few minutes, much faster than most existing

where Γ_k is a clustering that partitions the nodes in a graph into k groups, e_{ii} is the fraction of edges with both nodes within cluster i , and a_i is the fraction of edges with one or both nodes in cluster i . Intuitively, the Q function measures the percentage of edges fully contained within the clusters, subtracted by what one would expect if the edges were randomly placed. The value of Q is between -1 and 1; a larger Q value means stronger modular structures. If a partition gives no more

within-cluster edges than expected by chance, $Q \leq 0$. For a trivial partitioning with a single cluster, $Q = 0$. It has been observed that most real-world networks have $Q > 0.3$ [26]. The Q function can also be extended to weighted networks straightforwardly by generalizing e_{ii} and a_i to edge weights, instead of number of edges.

Since the optimization of Q is NP-hard [52,53], we developed a heuristic procedure to achieve this goal. Given the adjacency matrix of a network G , we apply a standard spectral clustering algorithm [45] to search for the best 2-, 3, or 4-way partitioning that gives the highest Q value. This is recursively applied to partition each subnetwork until the overall Q value of the network does not increase. To further optimize Q , an efficient greedy search procedure is then repeatedly applied to look for the following possible operations: merging two modules, moving a node from one module to a different module, or further splitting a module. The procedure terminates when no operations can improve Q .

Module evaluation

Comparing to known modular structures

To compare the modules identified by an algorithm to the true modules, we computed the adjusted Rand Index [40]. Given a set of objects $S = s_1, s_2, \dots, s_m$, let $X = \{X_1, X_2, \dots, X_M\}$ and $Y = \{Y_1, Y_2, \dots, Y_N\}$ represent the true and predicted partitions of the objects, where each object appears in X and Y exactly once. Let n_{ij} be the number of common objects between X_i and Y_j . Also let $n_{i\bullet} = \sum_j n_{ij} = |X_i|$ be the size of X_i , and $n_{\bullet j} = \sum_i n_{ij} = |Y_j|$ be the size of Y_j . The adjusted Rand Index can be computed by:

$$R(X, Y) = \frac{\sum_{i,j} \binom{n_{ij}}{2} - \sum_i \binom{n_{i\bullet}}{2} \sum_j \binom{n_{\bullet j}}{2} / \binom{n}{2}}{\frac{1}{2} \left[\sum_i \binom{n_{i\bullet}}{2} + \sum_j \binom{n_{\bullet j}}{2} \right] - \sum_i \binom{n_{i\bullet}}{2} \sum_j \binom{n_{\bullet j}}{2} / \binom{n}{2}} \quad (2)$$

Statistical enrichment of GO terms and transcription factor targets

To assess the functional significance of gene modules, we first compute the enrichment of GO terms for the genes within each module. The statistical significance of GO term enrichment is measured by a cumulative hypergeometric test [54]. The p -values are adjusted by Bonferroni corrections for the multiple testing problem [54]. The search of enriched GO terms is performed with a computer program GO::TermFinder [55].

To compare different results with approximately the same number of modules, we count the number of GO terms enriched in the modules at a given significance level. Furthermore, to rule out the possibility that a single module may contain a very large number of enriched GO terms and therefore dominate the contribution from

other modules, we also compute the percentage of modules that have at least one enriched GO term at a given significance level. It is worth noting that two sets of results cannot be compared by this method if they differ significantly in numbers of modules or modules size distributions, which may strongly affect the number of enriched GO terms. The results of the comparison may also depend on what p -value threshold is used.

The enrichment of transcription factor targets was determined similarly as in computing the enrichment of GO terms, and the binding data were from the large-scale ChIP-chip assay of 203 yeast transcription factors (TFs) under rich media conditions [56]. We only consider a binding as real if its p -value is less than 0.001, according to the original authors [56].

Reference network-based module evaluation

We propose a novel method for assessing the functional significance of gene modules. The basic idea is to introduce a functional reference network (discussed later), and compare the gene modules in a co-expression network with the structures of the reference networks. In such a reference network, genes are linked by edges that represent certain functional relationships between them, where the edges may be weighted according to the reliability or significance of the relationships. This network can be expected to have some modular structures as well. Since our purpose is to identify functional modules within a co-expression network, we would prefer a good partitioning of the gene co-expression network to represent a good partitioning of the reference network as well; i.e., genes within the same co-expression module should be connected by many high weight edges in the reference network, while genes in different co-expression modules should share less functions or be connected with low weight edges in the reference network. To quantify this, we force the reference network to be partitioned exactly the same way as the co-expression network, i.e., the group memberships of the nodes in the reference network are the same as that of the co-expression network. We then score the gene modules by the modularity of the reference network using Equation (1). The modularity score is between -1 and 1, with one meaning the co-expression network modules perfectly agree with the modular structure of the reference network. Since this measure is not biased by the number of modules or the module size distributions, it can be applied to compare arbitrary clustering results.

A reference network can be obtained from a variety of sources. First, available biological networks, such as PPI networks and genetic interaction networks, can be used as reference networks directly. A reference network can also be derived from other attributes of genes. In general, two genes can be connected if they possess some common attributes, given that the common attributes

are related to co-expression. For example, co-expressed genes may participate in the same biological process or be regulated by a common TF. These types of information can be represented by a matrix, where each row is a gene, and each column is an attribute. To construct a reference network from the matrix, genes are treated as nodes, and an edge is drawn between two genes if they share at least one common attribute. Edges are weighted by some similarity measure of genes' attributes. To measure the similarity, we use a well-known function in machine learning that takes into account the significance of attributes [57]. For example, the Gene Ontology term GO:0009987 (cellular process), which is very close to the root of the Gene Ontology graph and has a large number of genes associated, is less informative and should be weighted less than the term GO:0045911 (positive regulation of DNA recombination).

Denote a gene-attribute matrix by $A = (a_{ij})$, where $a_{ij} = 1$ if gene i has attribute j , or 0 otherwise. A is transformed into a weighted matrix $W = (w_{ij})$, where $w_{ij} = a_{ij} \times idf_j$. The weighting factor idf_j , called the inverse document frequency (IDF) [57], is defined by $idf_j = \log(n/\sum_i a_{ij})$, where n is the number of genes. With this transformation, the attributes that occur in many genes receive low weights in W . The edge weight between two genes is then measured by the cosine of their weighted attribute vectors:

$$S_{ij} = \cos(w_i, w_j) = \frac{\sum_k w_{ik} w_{jk}}{\sqrt{\sum_k w_{ik}^2 \sum_k w_{jk}^2}}, \quad (3)$$

where w_i and w_j are the i -th and j -th rows of W , respectively. As expected, many genes may be connected with very low weights if they share some non-specific functions. We apply a weight cutoff to remove such edges. We have found, however, that the result is almost not affected by the use of different cutoff values, as shown in Results and Discussion.

We use three types of reference networks to evaluate clusters. The first is a network constructed from Gene Ontology biological process terms [58], with each term as an attribute. The ontology and annotation files for yeast and Arabidopsis genes are downloaded from <http://www.geneontology.org/>. To construct a reference network, we first convert the original annotation files to include complete annotations, i.e., if a gene is associated with a certain term, we also add all ancestors of the term into the gene's attribute list due to term inheritance. If two terms are associated with exactly the same set of genes, we remove one to avoid double counting. We also remove GO terms that are associated with more than 500 or less than 5 genes. The procedure results in 1034 and 438 GO terms for yeast and

Arabidopsis, respectively. The second is a PPI network for budding yeast, downloaded from the BioGRID database [59]. We combined all physical interactions obtained from yeast two-hybrid or affinity purification-mass spectrometry experiments. The third network is a co-regulation network derived from the ChIP-chip data of 203 yeast transcription factors (TFs) under rich media conditions [56]. We treat each TF as an attribute, and construct a network with the procedure described above. We only consider a binding as real if its p -value is less than 0.001, according to the original authors [56].

Additional file 1: Figures S1-S3 and Tables S1-S3

Click here for file

[<http://www.biomedcentral.com/content/supplementary/1752-0509-4-8-S1.PDF>]

Acknowledgements

This work was supported in part by a UTSA faculty research award and a grant from the National Institute of General Medical Sciences (Award Number SC3GM086305) to J R, and NSF grants (IIS-0535257 and DBI-0743797) and NIH grants (RC1AR05868101 and U54AI05716006S1) to WZ. The content is solely the responsibility of the authors and does not necessarily represent the official views of the funding agencies. The authors wish to thank the anonymous reviewers for their constructive comments and suggestions.

Author details

¹Department of Computer Science, The University of Texas at San Antonio, One UTSA Circle, San Antonio, TX 78249, USA. ²Department of Computer Science and Engineering, Washington University in St Louis, One Brookings Dr, St Louis, MO 63130, USA. ³Department of Genetics, Washington University School of Medicine, Campus Box 8232, St Louis, MO 63110, USA.

Authors' contributions

JR and WZ conceived of the research and designed the study. JR carried out most of the computational analysis. AKD participated in evaluating the methods using synthetic microarray data. JR wrote the paper and WZ helped with the manuscript preparation. All authors read and approved the final manuscript.

Received: 27 July 2009

Accepted: 2 February 2010 Published: 2 February 2010

References

1. Eisen M, Spellman P, Brown P, Botstein D: **Cluster analysis and display of genome-wide expression patterns.** *Proc Natl Acad Sci USA* 1998, **95**:14863-8.
2. Tegner J, Yeung M, Hasty J, Collins J: **Reverse engineering gene networks: integrating genetic perturbations with dynamical modeling.** *Proc Natl Acad Sci USA* 2003, **100**:5944-9.
3. Friedman N, Linial M, Nachman I, Pe'er D: **Using Bayesian networks to analyze expression data.** *J Comput Biol* 2000, **7**:601-20.
4. Davidich M, Bornholdt S: **Boolean network model predicts cell cycle sequence of fission yeast.** *PLoS ONE* 2008, **3**:e1672.
5. Presson A, Sobel E, Papp J, Suarez C, Whistler T, Rajeevan M, Vernon S, Horvath S: **Integrated weighted gene co-expression network analysis with an application to chronic fatigue syndrome.** *BMC Syst Biol* 2008, **2**:95.
6. Ghazalpour A, Doss S, Zhang B, Wang S, Plaisier C, Castellanos R, Brozell A, Schadt E, Drake T, Lusis A, Horvath S: **Integrating genetic and network analysis to characterize genes related to mouse weight.** *PLoS Genet* 2006, **2**(8):e130.

7. Weston D, Gunter L, Rogers A, Wulfschleger S: **Connecting genes, coexpression modules, and molecular signatures to environmental stress phenotypes in plants.** *BMC Syst Biol* 2008, **2**:16.
8. Elo L, Jarvenpaa H, Oresic M, Lahtesmaa R, Aittokallio T: **Systematic construction of gene coexpression networks with applications to human T helper cell differentiation process.** *Bioinformatics* 2007, **23**(16):2096-103.
9. Oldham M, Horvath S, Geschwind D: **Conservation and evolution of gene coexpression networks in human and chimpanzee brains.** *Proc Natl Acad Sci USA* 2006, **103**(47):17973-8.
10. Lee H, Hsu A, Sajdak J, Qin J, Pavlidis P: **Coexpression analysis of human genes across many microarray data sets.** *Genome Res* 2004, **14**(6):1085-94.
11. Tsaparas P, Marino-Ramirez L, Bodenreider O, Koonin E, Jordan I: **Global similarity and local divergence in human and mouse gene co-expression networks.** *BMC Evol Biol* 2006, **6**:70.
12. Jordan I, Marino-Ramirez L, Wolf Y, Koonin E: **Conservation and coevolution in the scale-free human gene coexpression network.** *Mol Biol Evol* 2004, **21**(11):2058-70.
13. van Noort V, Snel B, Huynen M: **The yeast coexpression network has a small-world, scale-free architecture and can be explained by a simple model.** *EMBO Rep* 2004, **5**(3):280-4.
14. Zhu D, Hero A, Cheng H, Khanna R, Swaroop A: **Network constrained clustering for gene microarray data.** *Bioinformatics* 2005, **21**:4014-20.
15. Carter S, Brechbuhler C, Griffin M, Bond A: **Gene co-expression network topology provides a framework for molecular characterization of cellular state.** *Bioinformatics* 2004, **20**:2242-50.
16. Zhou X, Kao M, Wong W: **Transitive functional annotation by shortest-path analysis of gene expression data.** *Proc Natl Acad Sci USA* 2002, **99**:12783-8.
17. Aggarwal A, Guo D, Hoshida Y, Yuen S, Chu K, So S, Boussioutas A, Chen X, Bowtell D, Aburatani H, Leung S, Tan P: **Topological and functional discovery in a gene coexpression meta-network of gastric cancer.** *Cancer Res* 2006, **66**:232-41.
18. Stuart J, Segal E, Koller D, Kim S: **A gene-coexpression network for global discovery of conserved genetic modules.** *Science* 2003, **302**(5643):249-55.
19. Faith JJ, Hayete B, Thaden JT, Mogno I, Wierzbowski J, Cottarel G, Kasif S, Collins JJ, Gardner TS: **Large-Scale mapping and validation of Escherichia coli transcriptional regulation from a compendium of expression profiles.** *PLoS Biol* 2007, **5**:e8.
20. Ray M, Ruan J, Zhang W: **Variations in the transcriptome of Alzheimer's disease reveal modular networks involved in cardiovascular diseases.** *Genome Biology* 2008, **9**:R148.
21. Horvath S, Dong J: **Geometric interpretation of gene coexpression network analysis.** *PLoS Comput Biol* 2008, **4**(8):e1000117.
22. Srinivasanagendra V, Page G, Mehta T, Coulibaly I, Loraine A: **CressExpress: a tool for large-scale mining of expression data from Arabidopsis.** *Plant Physiol* 2008, **147**(3):1004-16.
23. Obayashi T, Hayashi S, Shibaoka M, Saeki M, Ohta H, Kinoshita K: **COXPRESdb: a database of coexpressed gene networks in mammals.** *Nucleic Acids Res* 2008, **36** Database: D77-82.
24. Magwene P, Kim J: **Estimating genomic coexpression networks using first-order conditional independence.** *Genome Biol* 2004, **5**(12):R100.
25. Carlson MR, Zhang B, Fang Z, Mischel PS, Horvath S, Nelson SF: **Gene connectivity, function, and sequence conservation: predictions from modular yeast co-expression networks.** *BMC Genomics* 2006, **7**:40.
26. Newman M: **The structure and function of complex networks.** *SIAM Review* 2003, **45**:167-256.
27. Barabasi A, Oltvai Z: **Network biology: understanding the cell's functional organization.** *Nat Rev Genet* 2004, **5**:101-13.
28. Ruan J, Zhang W: **Identifying network community structures with a high resolution.** *Phys Rev E Stat Nonlin Soft Matter Phys* 2008, **77**:016104.
29. Dongen SV: **Graph clustering via a discrete uncoupling process.** *SIAM Journal on Matrix Analysis and Applications* 2008, **30**:121-41.
30. Alizadeh A, Eisen M, Davis R, Ma C, Lossos I, Rosenwald A, Boldrick J, Sabet H, Tran T, Yu X, Powell J, Yang L, Marti G, Moore T, J JH, Lu L, Lewis D, Tibshirani R, Sherlock G, Chan W, Greiner T, Weisenburger D, Armitage J, Warnke R, Levy R, Wilson W, Grever M, Byrd J, Botstein D, Brown P, Staudt L: **Distinct types of diffuse large B-cell lymphoma identified by gene expression profiling.** *Nature* 2000, **403**:503-11.
31. Cooper C, Frieze AM: **On the connectivity of random k-th nearest neighbour graphs.** *Combinatorics, Probability & Computing* 1995, **4**:343-62.
32. Jeong H, Mason S, Barabasi A, Oltvai Z: **Lethality and centrality in protein networks.** *Nature* 2001, **411**:41-2.
33. Ravasz E, Somera A, Mongru D, Oltvai Z, Barabasi A: **Hierarchical organization of modularity in metabolic networks.** *Science* 2002, **297**:1551-5.
34. Albert R, Barabasi A: **Statistical mechanics of complex networks.** *Reviews of Modern Physics* 2002, **74**:47.
35. Keller E: **Revisiting scale-free networks.** *BioEssays* 2005, **27**:1060-8.
36. Siegal M, Promislow D, Bergman A: **Functional and evolutionary inference in gene networks: does topology matter?.** *Genetica* 2007, **129**:83-103.
37. Gasch A, Spellman P, Kao C, Carmel-Harel O, Eisen M, Storz G, Botstein D, Brown P: **Genomic expression programs in the response of yeast cells to environmental changes.** *Mol Biol Cell* 2000, **11**:4241-57.
38. Fjallstrom P: **Algorithms for graph partitioning: A survey.** *Linkoping Electron Atricles in Comput and Inform Sci* 1998.
39. Thalamuthu A, Mukhopadhyay I, Zheng X, Tseng G: **Evaluation and comparison of gene clustering methods in microarray analysis.** *Bioinformatics* 2006, **22**(19):2405-12.
40. Meila M: **Comparing clusterings-an information based distance.** *Journal of Multivariate Analysis* 2007, **98**(5):873-95.
41. Tibshirani R, Walther G, Hastie T: **Estimating the number of clusters in a data set via the gap statistic.** *Journal Of The Royal Statistical Society Series B* 2001, **63**:411-23.
42. Rousseeuw P: **Silhouettes: a graphical aid to the interpretation and validation of cluster analysis.** *J Comput Appl Math* 1987, **20**:53-65.
43. Azuaje F: **A cluster validity framework for genome expression data.** *Bioinformatics* 2002, **18**(2):319-20.
44. Tamayo P, Slonim D, Mesirov J, Zhu Q, Kitarawan S, Dmitrovsky E, Lander E, Golub T: **Interpreting patterns of gene expression with self-organizing maps: methods and application to hematopoietic differentiation.** *Proc Natl Acad Sci USA* 1999, **96**:2907-12.
45. Ng AY, Jordan MI, Weiss Y: **On spectral clustering: analysis and an algorithm.** *NIPS* 2001, **849**-56.
46. Herrero J, Valencia A, Dopazo J: **A hierarchical unsupervised growing neural network for clustering gene expression patterns.** *Bioinformatics* 2001, **17**(2):126-36.
47. Dwight S, Balakrishnan R, Christie K, Costanzo M, Dolinski K, Engel S, Feierbach B, Fisk D, Hirschman J, Hong E, Issel-Tarver L, Nash R, Sethuraman A, Starr B, Theesfeld C, Andrada R, Binkley G, Dong Q, Lane C, Schroeder M, Weng S, Botstein D, Cherry J: **Saccharomyces genome database: underlying principles and organisation.** *Brief Bioinform* 2004, **5**:9-22.
48. Ellis D, Eaton M, Fox R, Juneja S, Leong A, Miliuskas J, Norris D, Spagnolo D, Turner J: **Diagnostic pathology of lymphoproliferative disorders.** *Pathology* 2005, **37**:434-56.
49. Jaeger J, Koczan D, Thiesen H, Ibrahim S, Gross G, Spang R, Kunz M: **Gene expression signatures for tumor progression, tumor subtype, and tumor thickness in laser-microdissected melanoma tissues.** *Clin Cancer Res* 2007, **13**:806-15.
50. Shipp M, Ross K, Tamayo P, Weng A, Kutok J, Aguiar R, Gaasenbeek M, Angelo M, Reich M, Pinkus G, Ray T, Koval M, Last K, Norton A, Lister T, Mesirov J, Neuberg D, Lander E, Aster J, Golub T: **Diffuse large B-cell lymphoma outcome prediction by gene-expression profiling and supervised machine learning.** *Nat Med* 2002, **8**:68-74.
51. Newman M, Girvan M: **Finding and evaluating community structure in networks.** *Phys Rev E Stat Nonlin Soft Matter Phys* 2004, **69**:026113.
52. Garey M, Johnson D: **Computers and Intractability: A Guide to the Theory of NP-completeness.** *Freeman, San Francisco* 1979.
53. Brandes U, Delling D, Höfer M, Gaertler M, Görke R, Nikoloski Z, Wagner D: **On finding graph clusterings with maximum modularity.** *Proceedings of the 33rd International Workshop on Graph-Theoretic Concepts in Computer Science (WG'07) Lecture Notes in Computer Science*, Springer 2007.
54. Altman D: **Practical Statistics for Medical Research** Chapman & Hall/CRC, London 1991.
55. Boyle E, Weng S, Gollub J, Jin H, Botstein D, Cherry J, Sherlock G: **GO::TermFinder - open source software for accessing Gene Ontology information and finding significantly enriched Gene Ontology terms associated with a list of genes.** *Bioinformatics* 2004, **20**:3710-5.
56. Harbison C, Gordon D, Lee T, Rinaldi N, Macisac K, Danford T, Hannett N, Tagne J, Reynolds D, Yoo J, Jennings E, Zeitlinger J, Pokholok D, Kellis M, Rolfe P, Takusagawa K, Lander E, Gifford D, Fraenkel E, Young R:

Transcriptional regulatory code of a eukaryotic genome. *Nature* 2004, **431**:99-104.

57. Jones KS: IDF term weighting and IR research lessons. *Journal of Documentation* 2004, **60**:521-3.
58. Gene Ontology Consortium: The Gene Ontology (GO) database and informatics resource. *Nucleic Acids Res* 2004, **32** Database: D258-61.
59. Stark C, Breitkreutz B, Reguly T, Boucher L, Breitkreutz A, Tyers M: BioGRID: a general repository for interaction datasets. *Nucleic Acids Res* 2006, **34**: D535-9.

doi:10.1186/1752-0509-4-8

Cite this article as: Ruan *et al.*: A general co-expression network-based approach to gene expression analysis: comparison and applications. *BMC Systems Biology* 2010 **4**:8.

**Submit your next manuscript to BioMed Central
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

